TWO NEW SPECIES AND 
ONE SUBSPECIES OF RIODINID 
FROM SOUTHWEST BRAZIL 
(LEPIDOPTERA: RIODINIDAE) 

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ABSTRACT. — Two new species and one subspecies in the riodinid genera Xenandra C. & R. Felder, 1865, Argyrogrammana Strand, 1932, and Pachythone Bates, 1868, are described from Mato Grosso and Rondônia states in south-west Brazil. The symmachine species caerulea Godman & Salvin, 1878, is transferred to the genus Esthemopsis C. & R. Felder, 1865, from Xenandra (stat. rev.).

KEY WORDS: Amazon, antroconia, Argyrogrammana, Argyrogrammana talboti naranjilla n. sp., Bolivia, Dioptinae, Ecuador, Esthemopsis, Guianas, Leguminosae, Mato Grosso, Mesena, Mesopena, Neotropical, Pachythone, Pachythone anaculinus n. sp., Peru, Proteacaeceae, Rondônia, Sapindaceae, Steltiella, Symmachini, taxonomy, Xenandra, Xenandra mielkei n. sp.

The purpose of this paper is to describe three new riodinid taxa in the genera Xenandra C. & R. Felder, 1865, Argyrogrammana Strand, 1932, and Pachythone Bates, 1868, to facilitate the compilation of butterfly checklists for the Alto Rio Arinos area of Diamantino in Mato Grosso state, Brazil (EF), and Ecuador (JPWH). P. J. DeVries, as the co-discoverer of the new Pachythone species, is included as an author on that taxon. All of the unstarred collections listed in Hall (1999) have been examined for relevant types and additional material of the taxa described here. The following collection acronyms are used throughout the text:

AMNH American Museum of Natural History, New York, USA
BMNH (British) Natural History Museum, London, England
EF Eurides Furtado collection, Diamantino, Mato Grosso, Brazil
PHD Philip J. DeVries collection, Eugene, Oregon, USA
UFPC Universidade Federal do Parã£nd, Departamento de Zoologia, Curitiba, Parã£nd, Brazil
USNM United States National Museum, Smithsonian Institution, Washington, USA

Xenandra mielkei Hall & Furtado, n. sp. 

Fig. 1a-d; 4a-c

Description. — Male: Forewing length 15.5 mm. Forewing costal margin shallowly convex to base, distal margin strongly convex; hindwing angular, tornus pointed. Dorsal surface: forewing ground color dark brown; dark orange-red at very base of anal margin and along vein at lower edge of discal cell, becoming slightly broader in postdiscal region and extending to base of veins Cu1 and Cu2 and along discal cell end; fringe brown. Hindwing ground color dark brown, pale brown at very base of wing; large patch of dark orange-red extends from costal margin to a point in middle of cell 2A two-fifths distance from base to wing margin, and then as a semicircle to apex, small area of dark orange-red scaling in lower middle portion of cell 2A; long orange-brown, erectile, androconial setae along medial region of cell 2A extend towards anal margin; fringe brown. Ventral surface: ground color of both wings dark brown, paler brown at anal margin of forewing; orange-red dorsal pattern very faintly visible. Head: first and third segments of labial palpi brown, dorsal surface of second segment brown, ventral surface orange; third segment very short. Eyes brown and bare, margins with orange scaling. Frons orange. Antennial segments and tubular clubs entirely black. Body: both surfaces of thorax black, patagia orange. Dorsal surface of abdomen black, sides and ventral surface of distal half orange, except for narrow medial black line. A broad band of concealed androconial scales with a small gap dorsally on upper half of abdominal tergites 4 and 5 (Fig. 4c) (see Harvey, 1987, and Hall & Willmott, 1996a, for SEM illustrations of this scale type). All legs brown. Genitalia (Fig. 4a,b); uncus rounded at lower posterior corner, produced into point at dorsal tip, tegumen triangular with large rectangular lightly sclerotised region anteriorly; falcob of medium size and width; vinculum a narrow ribbon, produced into broad, short sacculus ventrally; valvae consist of well sclerotised lower portion produced into a single rounded point, and a more lightly sclerotised upper portion encircling aedeagus and produced into a rounded lobe; aedeagus as even width and slightly downwardly pointed, anterior opening directed ventrally, posterior opening directed ventrally and to right with tip curled inwards; a single large cluster of hair-like cornuti; pedicel large and well sclerotised, produced into a pronouncedly rounded posterior curve.

Female: Forewing length 15.5 mm. Wing shape similar to that of male but both wings more narrow. Dorsal surface: forewing ground color brown; yellow longitudinal stripe extends and broadens from wing base to near distal margin, encompassing lower half of discal cell, upper basal half of cell 2A, and basal two-thirds of cells Cu1 to M1 (some brown scaling at upper edge of latter cell); fringe orange. Hindwing ground color brown; yellow longitudinal stripe extends and broadens from wing base to near distal margin, encompassing lower half of discal cell, upper basal half of cell 2A, basal half of cell Cu1, and basal three-quarters of cells Cu2 to M1 (some brown scaling at upper edge of latter cell); costal and distal fringes brown, anal fringe yellow. Ventral surface: differs from dorsal surface in following respects: yellow scaling present on forewing as a band along base of costa and as a small fleck in tornus, and on hindwing as two small yellow spots at wing base above discal cell. Head: labial palpi a mixture of yellow and brown scaling on all segments. Eyes brown and bare, margins with yellow scaling. Frons yellow. Antennal segments and tubular clubs entirely black. Body: thorax brown with yellow scaling posteriorly at sides on dorsal surface and as a spot at middle of sides on ventral surface; patagia brown. Dorsal surface of abdomen brown, remainder yellow except for narrow medial brown line at sides and on ventral surface. All legs yellow-brown.

Types. — Holotype: BRAZIL — Mato Grosso, Alto Rio Arinos, nr. Diamantino, 400m, 2 Nov 1989 (E. Furtado); to be deposited in the USNM.

Etymology. — This species is named in honor of our friend, the eminent Brazilian lepidopterist, Olaf H. H. Mielke.
Fig. 1-3. 1. *Xenandra mielkei* Hall & Furtado n. sp., holotype ♂ [USNM]: a) dorsal surface; b) ventral surface. Allotype ♀ [USNM]: c) dorsal surface; d) ventral surface. 2. *Argyrogrammana talboti talboti* Brévignon & Gallard, 1998, ♂ Alto Rio Arinos, nr. Diamantino, Brazil [USNM]: a) dorsal surface; b) ventral surface. Holotype ♀, "French Guiana" [BMNH]: c) dorsal surface; d) ventral surface. *Argyrogrammana talboti naranjilla* Hall & Furtado n. sp., holotype ♂ [AMNH]: c) dorsal surface; f) ventral surface. Allotype ♀ [USNM]: g) dorsal surface; h) ventral surface. 3. *Pachythone analuciae* Hall, Furtado & DeVries n. sp., holotype ♂ [USNM]: a) dorsal surface; b) ventral surface. Ecuadorian paratype ♂ [PJD]: c) dorsal surface; d) ventral surface.
Diagnosis. - Xenandra mielkei n. sp. presents a remarkably interesting phenotype that appears to be an evolutionary intermediate between the genera Xenandra and Mesenopsis Godman & Salvin, [1886], and its discovery provides evidence that Mesenopsis may be a derived clade within Xenandra. Longitudinal stripes represent a very rare wing pattern in butterflies and in the moths (primarily Dipsocera) which are believed to act as their mimetic models (Miller, 1996), thus it seems most likely that Mesenopsis species, whose sexes have both longitudinal stripes, are more derived than Xenandra species. In previously described species of the latter genus, the male is typically black with a variably sized red patch at the costal margin of the hindwing while the female has a transverse red forewing band. However, X. mielkei is unique in possessing a typical Xenandra-like male phenotype and a Mesenopsis-like female phenotype. It is most interesting to note the wing pattern variation among paratype males. The red hindwing patch varies in size and the extent to which it extends to the costal margin but, most importantly, the medial forewing red scaling varies from being completely absent (as in typical Xenandra males) to being very extensive and similar in shape to that of the female forewing yellow. It is not difficult to imagine how the hindwing red of the male could evolve into a longitudinal stripe by a reduction of red along the length of the costal margin. Unlike all other Xenandra species, X. mielkei lacks the apomorphic grayish-green interneural stripes on both ventral wings which perhaps also suggests that it may be close to the progenitor of the currently recognised genera Xenandra and Mesenopsis. Other external characters of X. mielkei that are typical of Mesenopsis include the orange/yellow patagia, frons and palpi.

Another character set indicating the close relationship between Xenandra and Mesenopsis is their distribution of concealed abdominal androconial scales in males. In the three Xenandra species examined and in all Mesenopsis species, there is a single band of these scales on segments 4 and 5. X. mielkei is typical in this respect except that there is a small gap dividing the band dorsally. It is important to note that the distribution pattern "1,1,1,1" on segments 4-7 in Mesenopsis pulchella Godman, 1903, reported by Harvey [1987] and subsequently cited by Hall & Willmott [1996a] is erroneous and should be "1,1" on segments 4 and 5. It is assumed that this entry was mixed by Harvey [1987] with Mesene margaretae (White, 1843), which was recorded as having a "1,1" pattern on segments 4 and 5, but which actually has a "1,1,1,1" pattern on segments 4-7 (note that the typical pattern for Mesene is "1,1,1" on segments 4-6). It is also worth noting here that the abdominal androconial pattern distribution, wing pattern and male genitalia of the species caerulea Godman & Salvin, 1878, which is currently treated in Xenandra (Bridge, 1994), place it in the genus Estheneopsis C. & R. Felder, [1865] (stat. rev.), as first suggested by Godman & Salvin [1886] and also recently by Hall & Willmott [1996a].

The male genitalia of X. mielkei exhibit some characters in common with members of Xenandra, some with members of Mesenopsis, and some that are unique. The shapes of the uncus and tegumen are most similar to those of Mesenopsis species, but the unusual lightly sclerotised rectangle at the anterior edge of the tegumen is only seen in Xenandra, in a much reduced form.

The valva, which are relatively small with a single posterior point and only a lightly sclerotised dorsal portion over the aedeagus, are most similar to those of Xenandra species, while the pronouncedly curved pedicel appears to be somewhat intermediate between that of Xenandra and Mesenopsis. In the examined Xenandra species the pedicel is simple and strap-like while in all Mesenopsis species the pedicel is highly modified into a posteriorly projecting horn (a recurring character state in the Symmachini; e.g. in the genera Stichella Zikán, 1949, and Mesene Doubleday, 1847). Finally, many characters of the aedeagus, including the ventral instead of anterior opening anteriorly, the ventral and right opening posteriorly, and the single cluster of long hair-like setae, do not occur in any of the Xenandra species examined and in no Mesenopsis species.

In conclusion, X. mielkei uniquely presents a blend of characters typical of the genera Xenandra and Mesenopsis, and the discussion above presents evidence that the latter genus may be phylogenetically embedded in the former or that X. mielkei may be closely related to the progenitor of both genera. Either way, it is highly probable that cladistic analyses of the Symmachini will show that these genera should be combined. With this outcome in mind, this new species is placed in the first described of the aforementioned genera, Xenandra.

Discussion. - X. mielkei is currently known only from the Diamantina region of central-western Mato Grosso state in south-west Brazil, although it will certainly be found to be more widespread in the south-western Amazon basin.

Both sexes of X. mielkei were encountered resting under leaves less than 2m above the ground along forest edges around mid-day, and females were also observed visiting flowers of Inga laurina (Leguminoseae) and a Roupala species (Proteaceae). Aegyrogrammata talothi naranjilla Hall & Furtado, n. sp.

Fig. 2c-h, 5a-b

Description. - MALE: forewing length 16.5mm. Forewing costa straight, distal margin very slightly convex; hindwing rounded. Dorsal surface: forewing ground color black; large orange-yellow patch extends from wing base to near submargin and an in-curving band around end of discal cell to near costa, with uneven distal margin, that is orange below vein Cu 4 and yellow in the upper distal corner of that cell and above, thin line of yellow scaling at distal margin of orange; three small, evenly spaced triangles in discal cell of which basal two orange, distal one yellow; two small pale blue squares in middle of cells M 3 and M 4 (very faint in paratype); thin, undulating, faintly discontinuous silver-blue submarginal line extends from tornus to apex, where elements are crescent-shaped and largest; fringe black with white scaling at margin of cells 2A, Cu 4, M 4, and M 5. Hindwing ground color black; large orange patch extends from wing base near submargin, distal margin uneven with thin line of yellow scaling, anal margin paler orange; thin, undulating, faintly discontinuous silver-blue submarginal line extends from tornus to apex; distal fringe black with white scaling at margin of cells Cu 4, M 4, anal and costal fringes orange. Ventral surface: differs from dorsal surface in following respects: blue squares in forewing cells M 3 and M 4 very faint, an additional more prominent one in cell M 4; apical element of forewing submarginal silver-blue line notably larger; two tiny black spots at upper base of hindwing discal cell. Head: labial palpi mixture of very long brown and cream setae. Eyes brown and bare, margins with orange scaling. Frons brown with some lateral cream scaling. Antennal segments (of paratype) black with basal white scaling, tubular clubs black. Body: dorsal surface of thorax black with extensive orange scaling; patagia orange. Abdomen orange with slightly paler cream-orange ventrally. All legs cream. Genitalia (Fig. 5a-b): uncus angular, slightly indented at dorsal tip; tegumen elongate, anteriorly notched; styles somewhat large; vinculum narrow ribbon, produced into long saccus ventrally; valvae narrow and elongate, small angular point at lower margin, narrow distal portion joined at tip by small, downwardly posteriorly pointing spine; aedeagus long, narrow and pointed at tip; long continuous sclerotised cornutal band with jagged dorsal margin; pedicel long, broad at middle supporting aedeagus.

FEMALE: differs from male in following respects: forewing length 21mm. Wing shape more elongate. Dorsal surface: all markings in forewing discal cell larger and orange, orange extends above vein Cu 4 at base of forewing postdisal band, blue squares in forewing subapex absent; line of yellow scaling at distal margin of orange on hindwing slightly broader. Ventral surface: same differences as dorsal surface except that orange does not extend above vein Cu 4 at base of forewing postdisal band and a small area of additional black scaling present towards upper base of cell 2A.

Types. - Holotype ♀ BRAZIL - "Amazonas"; in the AMNH.
Allootype ♀: BRAZIL. — Rondônia, Rio Jamari, nr. Aripoimas, 1500m, 7 Aug 1978 (E. Furtado); to be deposited in the USNM. Paratypes: 1 ♀: same data as AT; to be deposited in the USNM. 1 ♀: same data as AT; in the EF. 1 ♀: Amazonas, Rio Purus, Lábrea, Hyutana (Huitana), Nov 1913 (H. W. Wickham); in the BMNH.

Etymology. — This taxon is named after the Spanish word for "orange," with reference to its bright, unmarked orange coloration, which is so atypical for the genus.

Diagnosis. — The female of Argyrogrammama talboti Brévignon & Gallard, 1998, was erroneously associated with Talbot (1929) (and also D’Abera, 1994) with the male of the phenotypically quite distinct Argyrogrammama sticheli Talbot 1929, and the true male has remained unknown until now. There has even been doubt as to whether the allotype of A. sticheli (= A. talboti) belonged in Argyrogrammama (Brévignon & Gallard, 1995, 1998), but examination of the genitalia of male specimens, which are illustrated here for the first time, confirms that the species does in fact belong in that genus (see below). Both sexes of nominotypical A. talboti are figured here along with another phenotype which we describe as a subspecies of it. Since Argyrogrammama is a highly speciose genus whose species often differ by only very minor wing pattern characters (Brévignon & Gallard, 1995, 1998; Hall & Willmott, 1995, 1996b, 1998), the consistent wing pattern differences exhibited by A. talboti and A. naranjilla might actually constitute specific differences, but given the small number of specimens currently available and the allopatric distribution of the two phenotypes (see Fig. 7), we conservatively prefer to treat these two taxa as subspecies. Sympathy would be the ultimate proof of their specific status.

A. talboti naranjilla n. ssp. differs most notably from the nominate by having a sharply contrasted orange/yellow forewing patch with a thin line of yellow scaling around the distal margin of the orange on both wings. This difference is most pronounced in the females, for while the sexes of the new subspecies are monomorphic, the female of the nominate is somewhat paler than the male and a uniform yellow. Other consistent but more subtle differences include the presence in the new subspecies of two tiny black spots at the base of the discal cell on the ventral hindwing, a vertical or convex instead of markedly concave apical portion to the distal margin of the orange patch at the costa of the ventral hindwing, and the lack of orange scaling at the very distal margin of the hindwing. The male genitalia of the two taxa do not differ. The configuration of the male genitalia, particularly the shape of the uncus, pedicel and valvae, which are joined at their tip by a small posteriorly pointed hook, most closely resembles that of A. saphirina (Staudinger, 1887), which belongs in the somewhat very different “trochilina group” (sensu Hall & Willmott, 1996b). This perhaps suggests an evolutionary sequence for the ventral wing pattern of this genus from the plesiomorphic spotted condition in members of the “stilbe” and “amalfreda groups” (sensu Hall & Willmott, 1995, 1996b), to the banded pattern of “trochilina group” species, to the complete lack of markings in A. talboti. A. glaucopsis (Bates, 1868), A. praestigiosa (Stichel, 1929) and A. subota (Hewitson, 1877) certainly appear to represent evolutionary intermediates between the “amalfreda” and “trochilina groups”.

Discussion. — A. talboti naranjilla is currently known only from Rondônia and southwestern Amazonas states in Brazil but probably also occurs in Bolivia and southern Peru. H. Austin (pers. comm.) reports collecting an additional seven males of A. t. naranjilla in the vicinity of Cacaualândia, near the type locality in Rondônia (Jul-Nov). The nominate subspecies has a broader known range that extends from the Guianas (2 females examined from French Guiana) to the other side of the Amazon basin in Mato Grosso state, Brazil (2 males examined from the Alto Rio Arinos, nr. Diamantino) (see Fig. 7).
Pachyphone analuciae Hall, Furtado & DeVries, n. sp.

Fig. 3a-d; 6

Description.—MALE: forewing length 12mm. Wing shape compact. Forewing costa straight, distal margin slightly convex; hindwing rounded. Dorsal surface: forewing ground color brown; faint orange discal and postdiscal scaling, distal margin darker brown; fringe brown with white scaling at distal margin of cells 2A to M, most prominent in cells M3 and M5. Hindwing ground color brown; large orange patch occupies entire area below vein M, except very base of wing, all of cell M3 except small distal region, and basal half of cell M2; fringe brown. Ventral surface: forewing ground color brown, yellow-brown at anal margin, darker brown at distal margin, paler brown curving postdiscal area; three dark brown spots in discal cell, one at base, one at middle and one at cell end, two dark brown spots below discal cell, at upper base of cell 2A; one dark brown marginal spot in cells Cu1 to M2 and two in cell 2A lined distally and proximally with dirty cream crescents; become larger towards apex. Hindwing ground color brown; large orange patch as on dorsal surface; distribution of basal dark brown spots as on ventral forewing; one small dark brown marginal semicircle embedded within orange in cells 3A to Cu1, one dark brown spot in cells M1 to M4 lined distally and proximally with dirty cream crescents. Head: first and second segments of labial palpi brown, third segment dark brown. Eyes brown and bare, margins with brown scaling. Frons dark brown in dorsal two-thirds, brown in ventral third. Antennae very short, segments black with basal white scaling; clubs black and flattened, tips orange-brown. Body: thorax brown on both surfaces, abdomen orange-brown on dorsal surface cream-brown on ventral surface. All legs brown. Genitalia (Fig. 6): ventral portion of uncus rounded, dorsal portion shallowly indented; tegumen triangular; falcifer of medium size and width; vinculum narrow and ribbon-like, produced into successus that is horizontally broad and short, and ventrally somewhat elongate; valvae produced into a conical point, basal lateral bulge prominent; aedeagus short and of even width, posterior opening directed ventrally and to right, no cornuti present; pedicel broad at base.

FEMALE: unknown.

Types.—Holotype 4: BRAZIL—Mato Grosso, Alto Rio Arinos, nr. Diamantino, 400m, 29 Aug 1993 (E. Furtado); to be deposited in the USNM. Paratypes: 1♂: same locality data as HT, 12 Nov 1992 (E. Furtado); in the EF. 1♂: same locality data as HT, 29 Oct 1998 (E. Furtado); to be deposited in the UPPC. ECUADOR—1♂: Sucumbios, Rio Napo, Garzacocha, La Selva, 200m, 25 Apr 1993 (P. J. DeVries); in the PJF.

Etymology.—This species is named in honor of Ana Lucia Mazzotti, wife of Furtado.

Diagnosis.—The short, broadly clubbed antennae, squat abdomen and configurations of the male genitalia clearly place P. analuciae n. sp. in the genus Pachyphone. P. analuciae resembles only Pachyphone distigma Bates, 1868, but has the hindwing patch orange instead of red and much larger in extent, extending beyond the discal cell and to the anal and distal margins. Orange-brown scaling is also present on the dorsal surface of the abdomen in P. analuciae. The male genitalia of all examined Pachyphone species are very homogeneous and provide few characters for identification; those of P. analuciae and P. distigma do not differ.

Discussion.—P. analuciae is currently known only from the disjunct localities of east Ecuador and south-west Brazil, but these suggest a broadly Amazonian distribution for the species. P. distigma, perhaps its closest relative, has a similarly broad, partially overlapping, Amazonian range that extends from the base of the eastern Andes (Hall and Willmott, unpubl. data) to the Atlantic (TL: Pará). Pachyphone species are generally highly localised in time and space, and thus very rarely encountered (Bates, 1868; Brown, 1993; Brévignon and Gallard, 1998; pers. observ.). The recent discovery of P. analuciae and the fact that the sole Ecuadorian male was found dead on the forest floor, suggests that this species is no exception. The Brazilian males were encountered in a humid area of Cerrado resting beneath leaves within 2m of the ground; one of these males was found imbuing fluids from an extraloral nectary on the young shoots of a Magonia pubescens plant (Sapindaceae).

Fig. 7. A map of South America illustrating the distribution of both subspecies of Argyrogramma talhoi.

Many Pachyphone species, most notably P. conspersa Stichel, 1926, P. xanthe Bates, 1868, and P. lateritia Bates, 1868, are highly phenotypically variable both between and within populations. The Ecuadorian paratype (Fig. 3c,d) is a darker specimen with a black ground color and no faint orange scaling on the dorsal forewing. A thin black line or small spots are variably present at the distal margin of the orange on the dorsal hindwing.

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